

Realistic 3D Cell Modeling for FEM Simulation

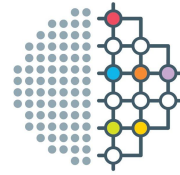
John Rugis
University of Auckland



*Workshop on Mathematical and Computational Methods in Biomedical Imaging and Image Analysis
University of Auckland, November 24, 2015*

Interdisciplinary research group

John Rugis
Computer Science



NeSI
New Zealand eScience
Infrastructure

James Sneyd
Shawn Means
Mathematics



THE UNIVERSITY
OF AUCKLAND
NEW ZEALAND

David Yule
Physiology



UNIVERSITY of
ROCHESTER

Project funding

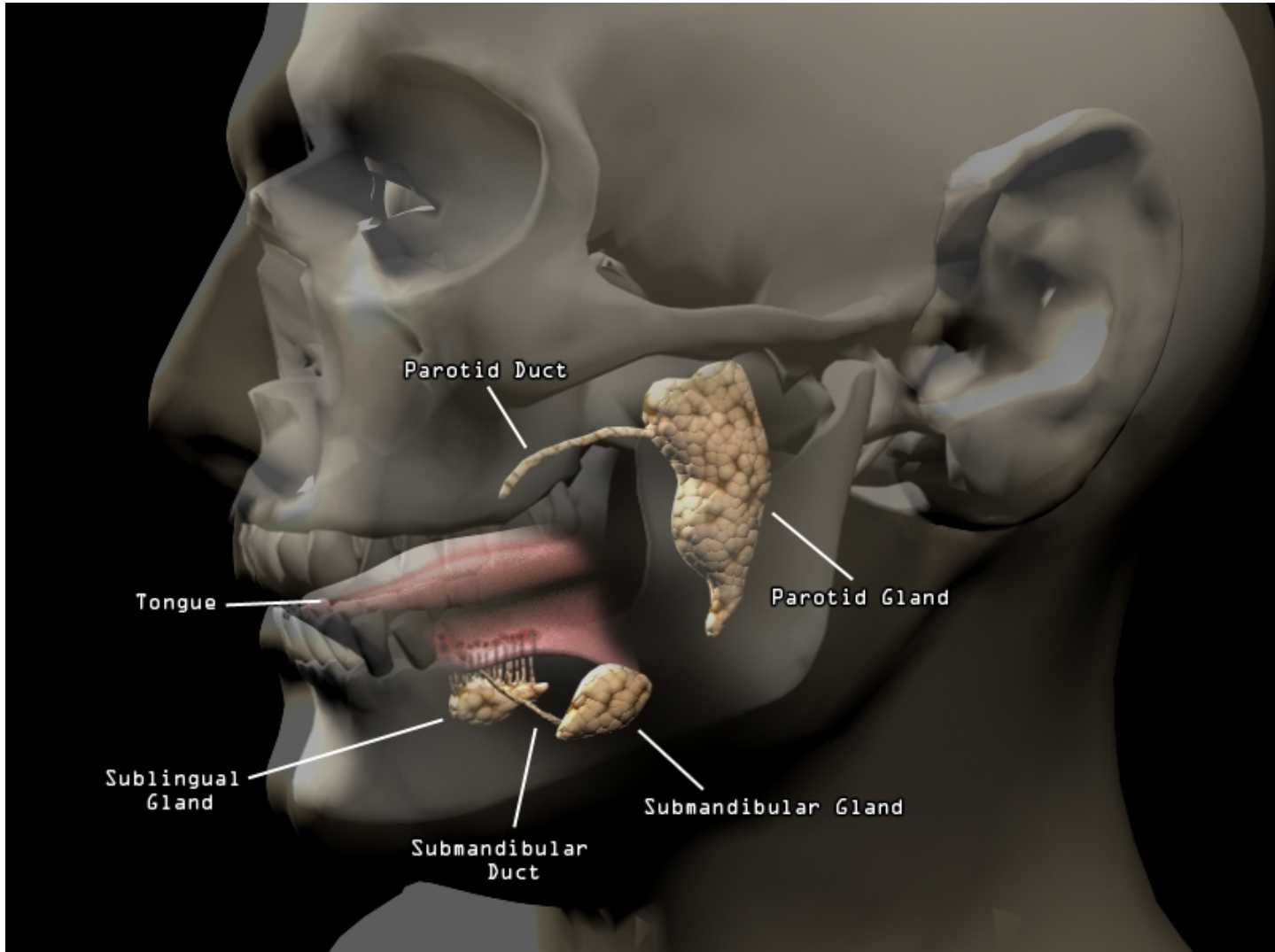


National Institutes of Health
Turning Discovery Into Health

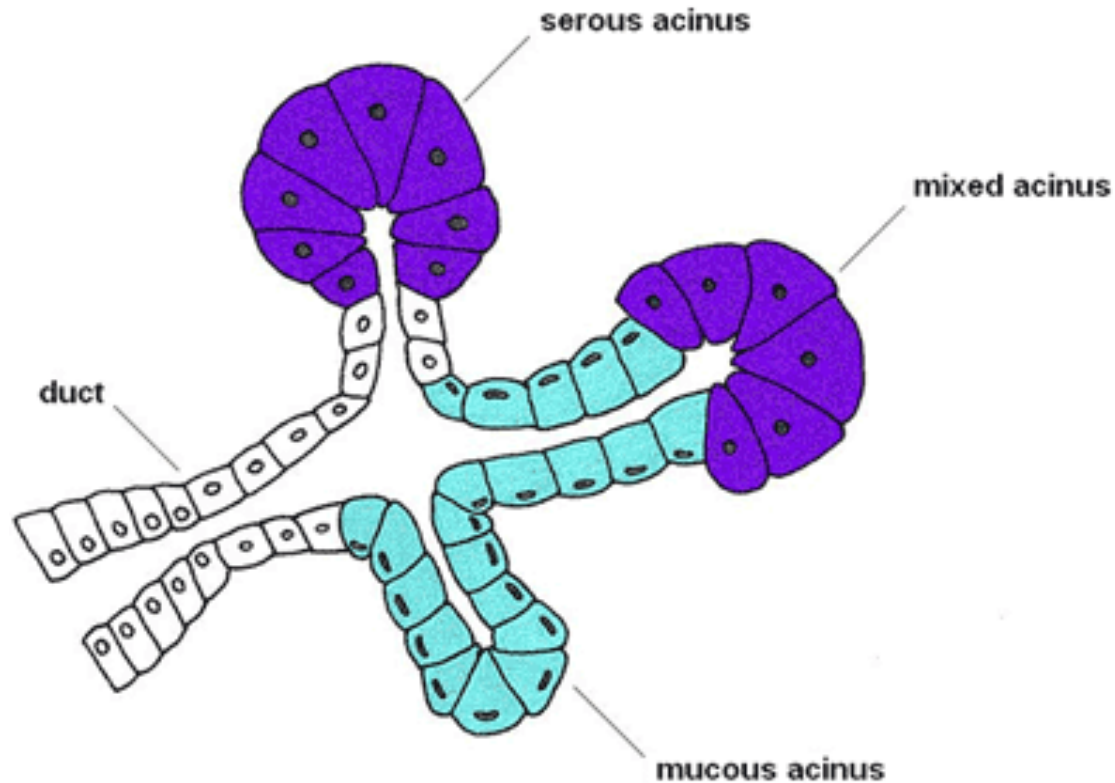
The goal:

Physically accurate modeling,
simulation and visualisation
of biological cell function.

Salivary glands



Salivary acinii



- **Three types:** serous, mucous and mixed

Data Acquisition: Confocal microscopy

2D image stacks

32 slices in Z direction.

Cells & Lumen

red: Na-KATPase

green: Cl Channel

Real dimensions used:
70.7 μm^2 , 2.2 μm spacing

TIFF files: 1024x1024



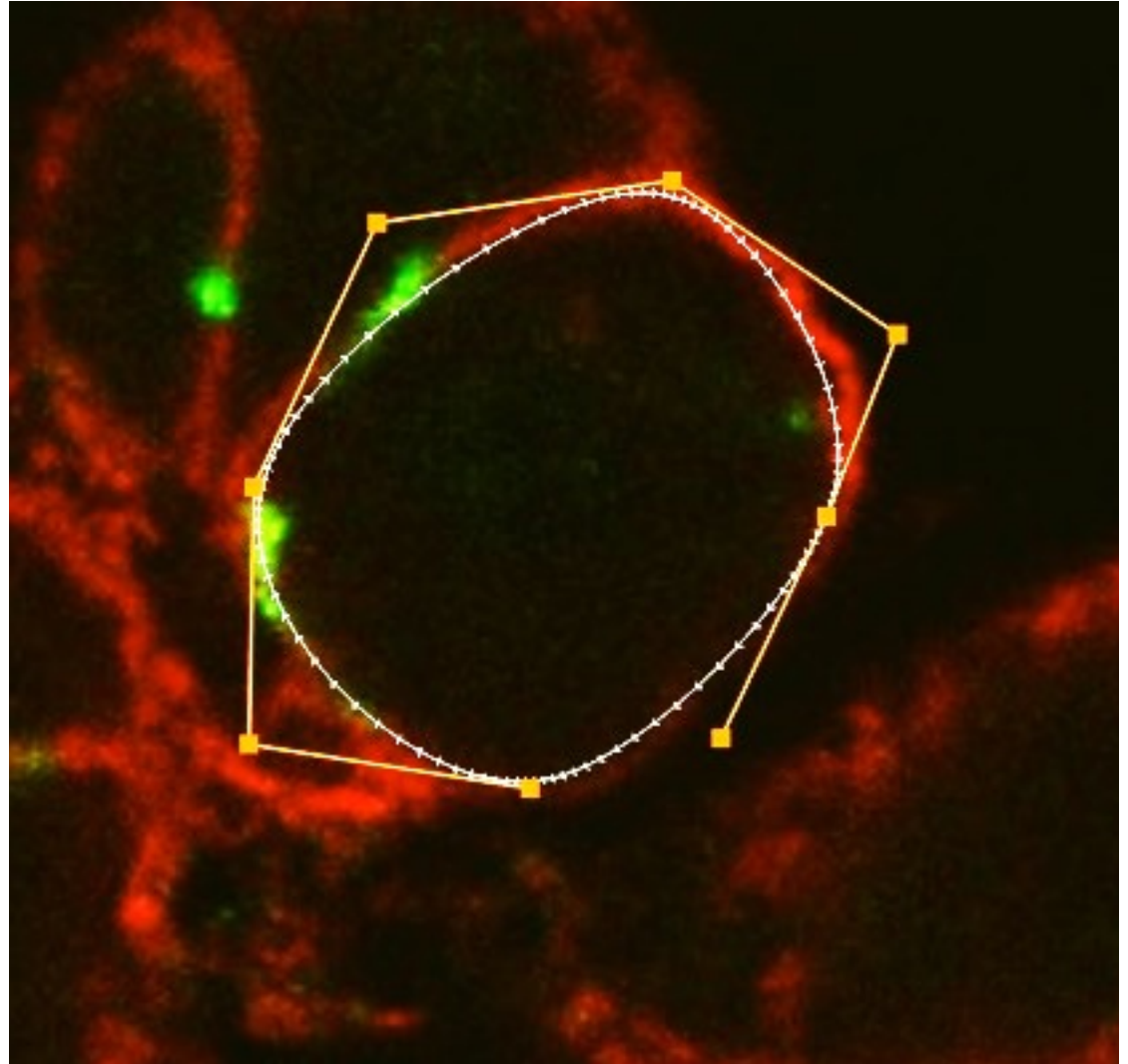
Mesh Modeling

3D reconstruction

Cells

1) Trace the outline of each cell in the image stack.

NURBS circles.



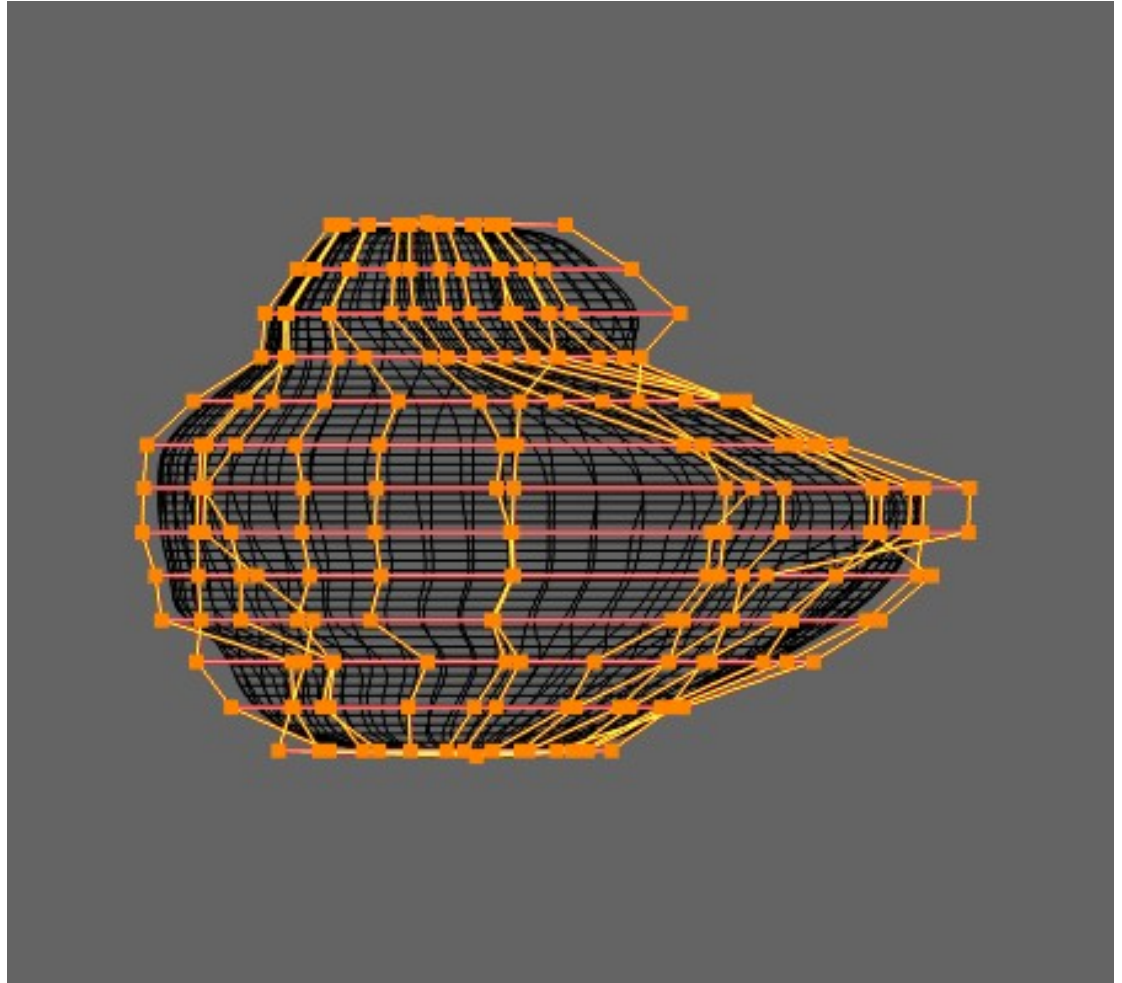
Mesh Modeling

3D reconstruction

Cells

2) Interpolate connections between the “level-set” outlines for each cell.

Results in a parametric NURBS surface.



Mesh Modeling

3D reconstruction

Cells

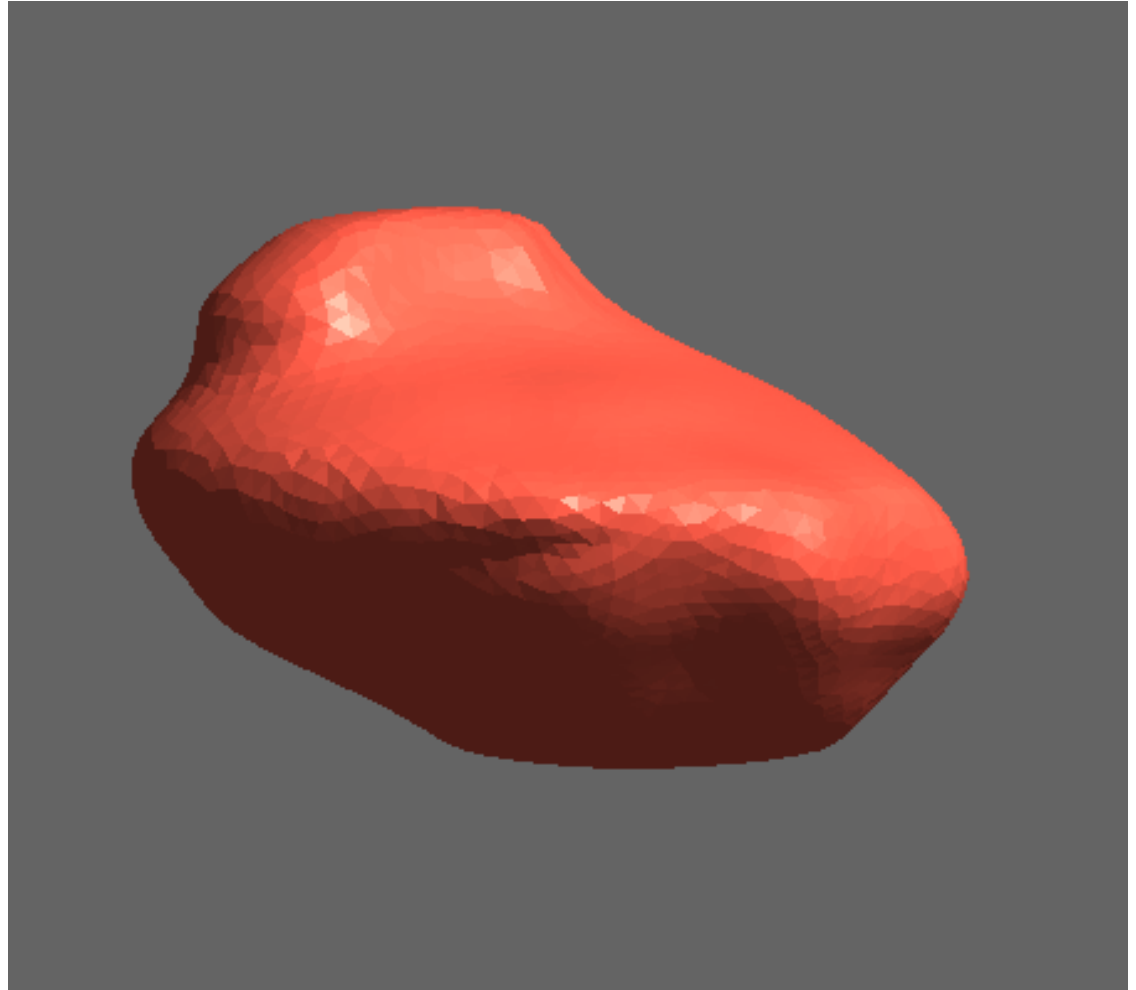
For FEM:

Refine the mesh
surface triangulation.

Octree remesh.

STL files.

Harmonic maps.



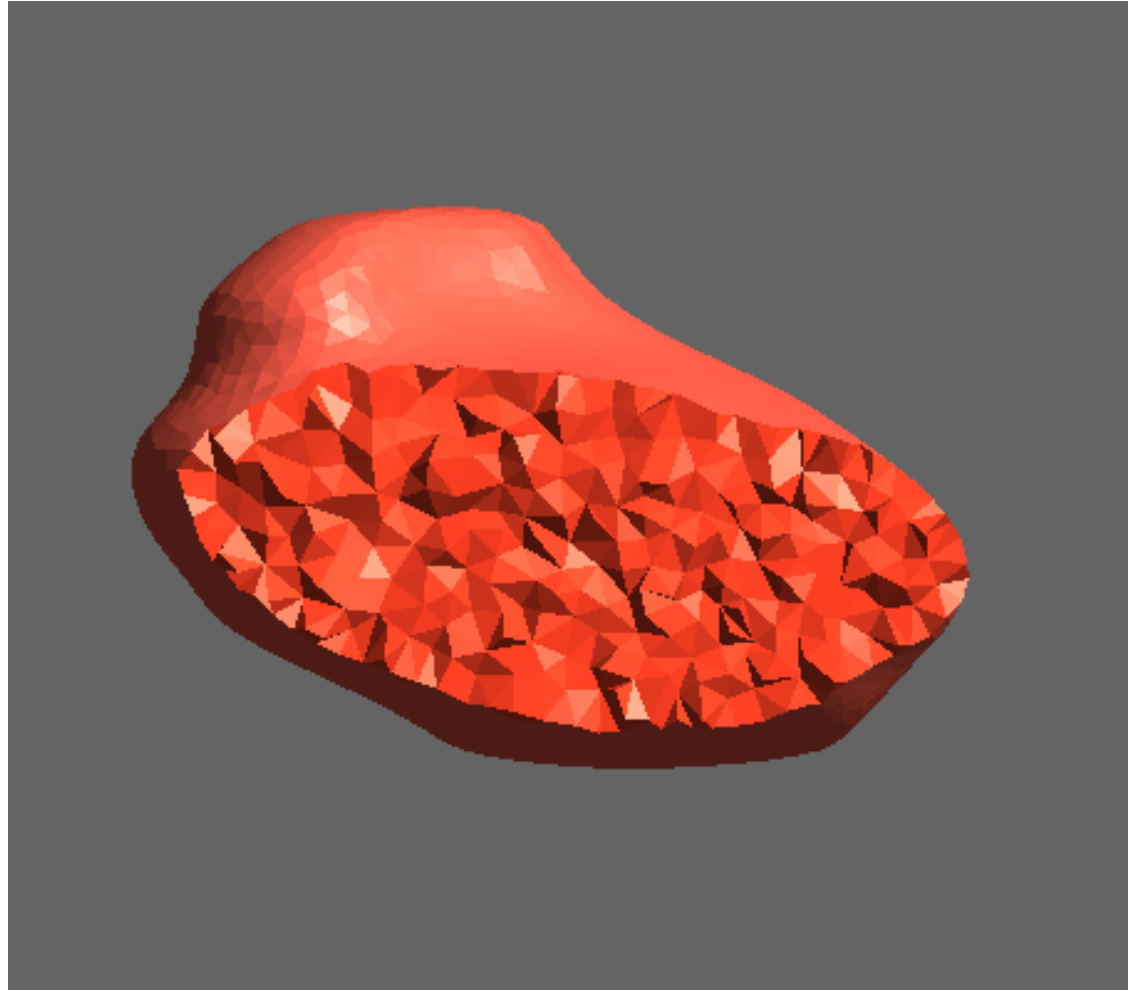
Mesh Modeling

3D reconstruction

Cells

For FEM:
Volumetric meshing
with tetrahedrons.

Harmonic maps.
MSH files.

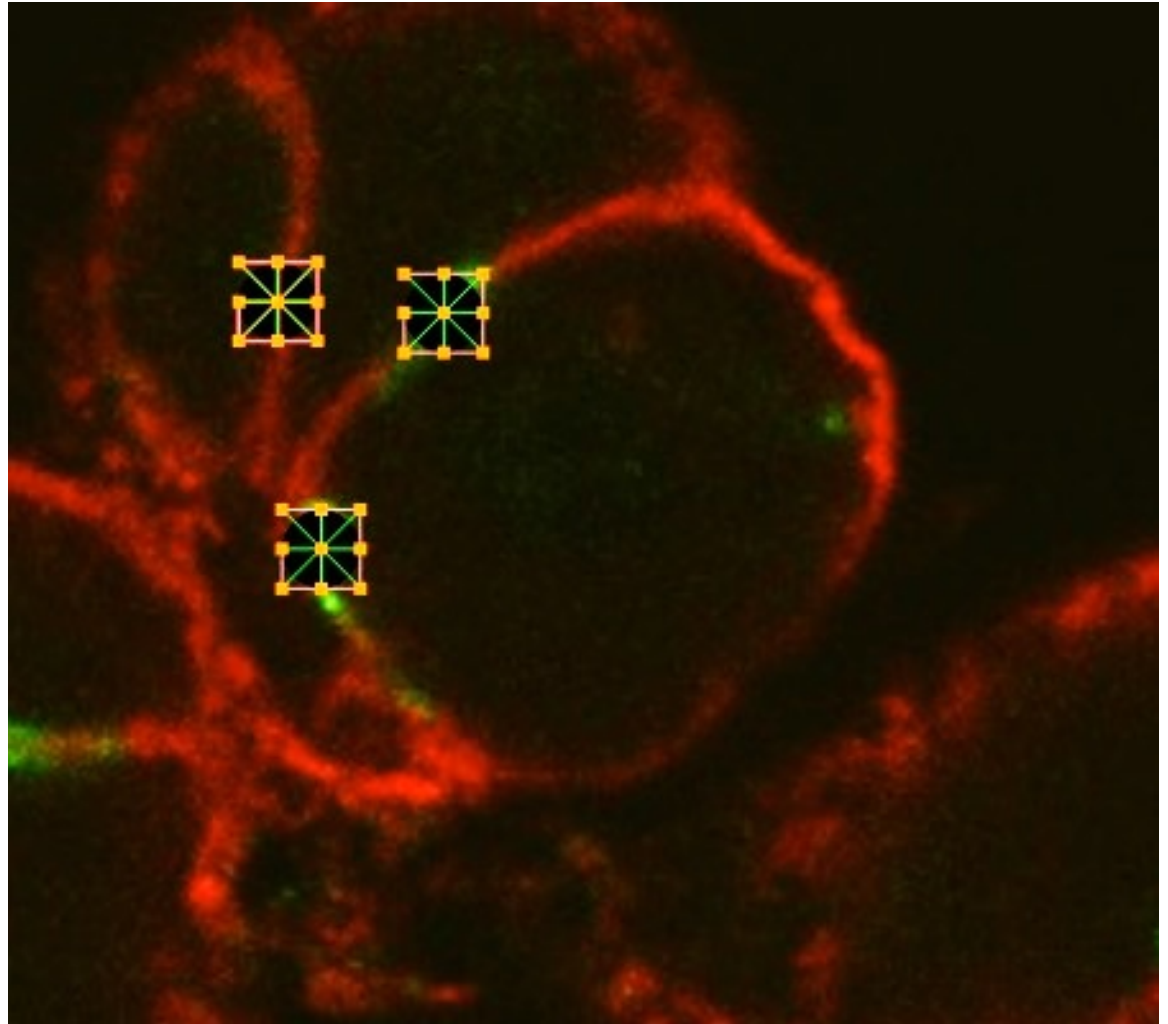


Mesh Modeling

3D reconstruction

Lumen

- 1) Place spheres on the lumen in each 2D image slice.



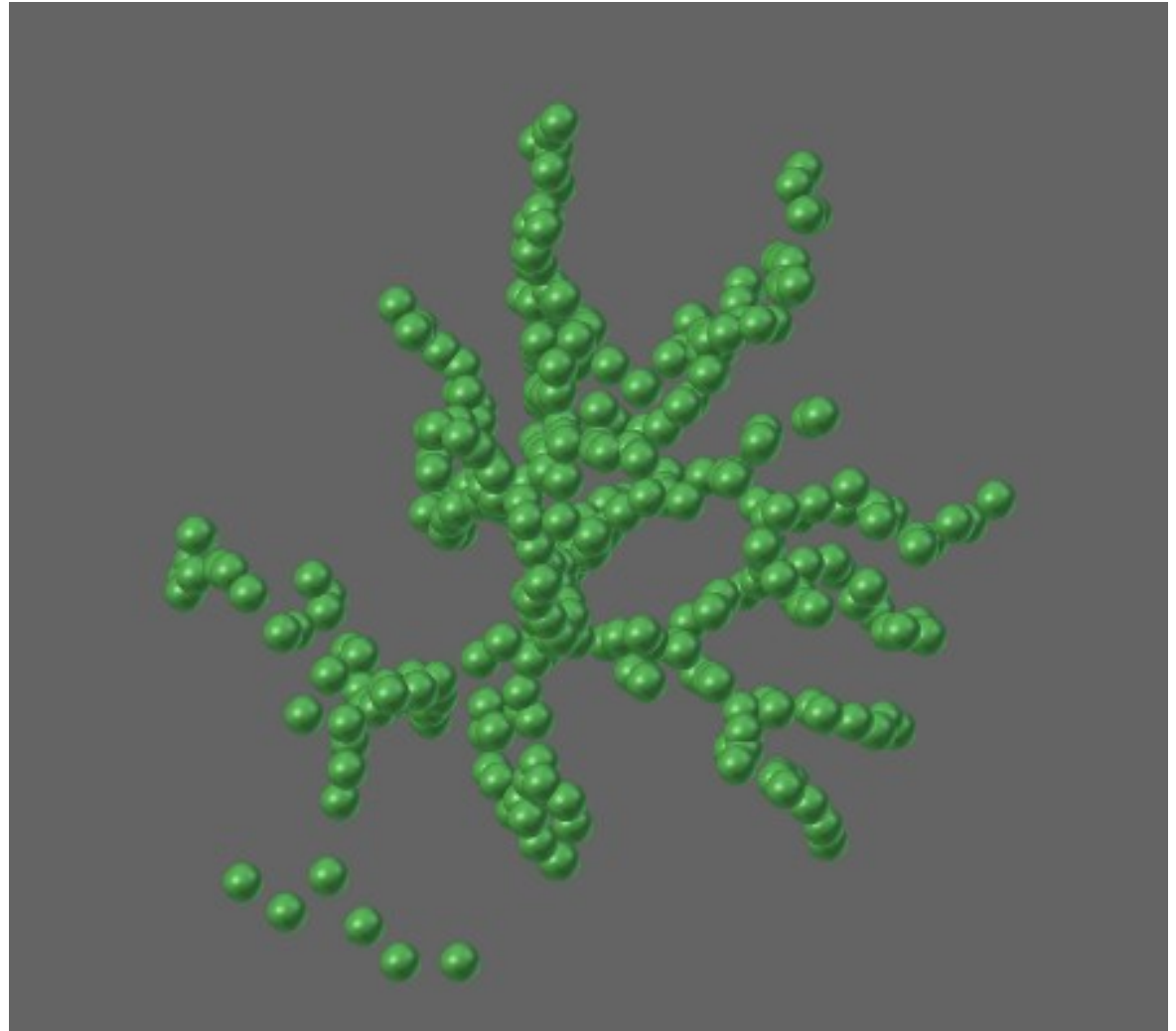
Mesh Modeling

3D reconstruction

Lumen

2) Collect all of the spheres together in 3D space.

Procedural “meta-balls”.



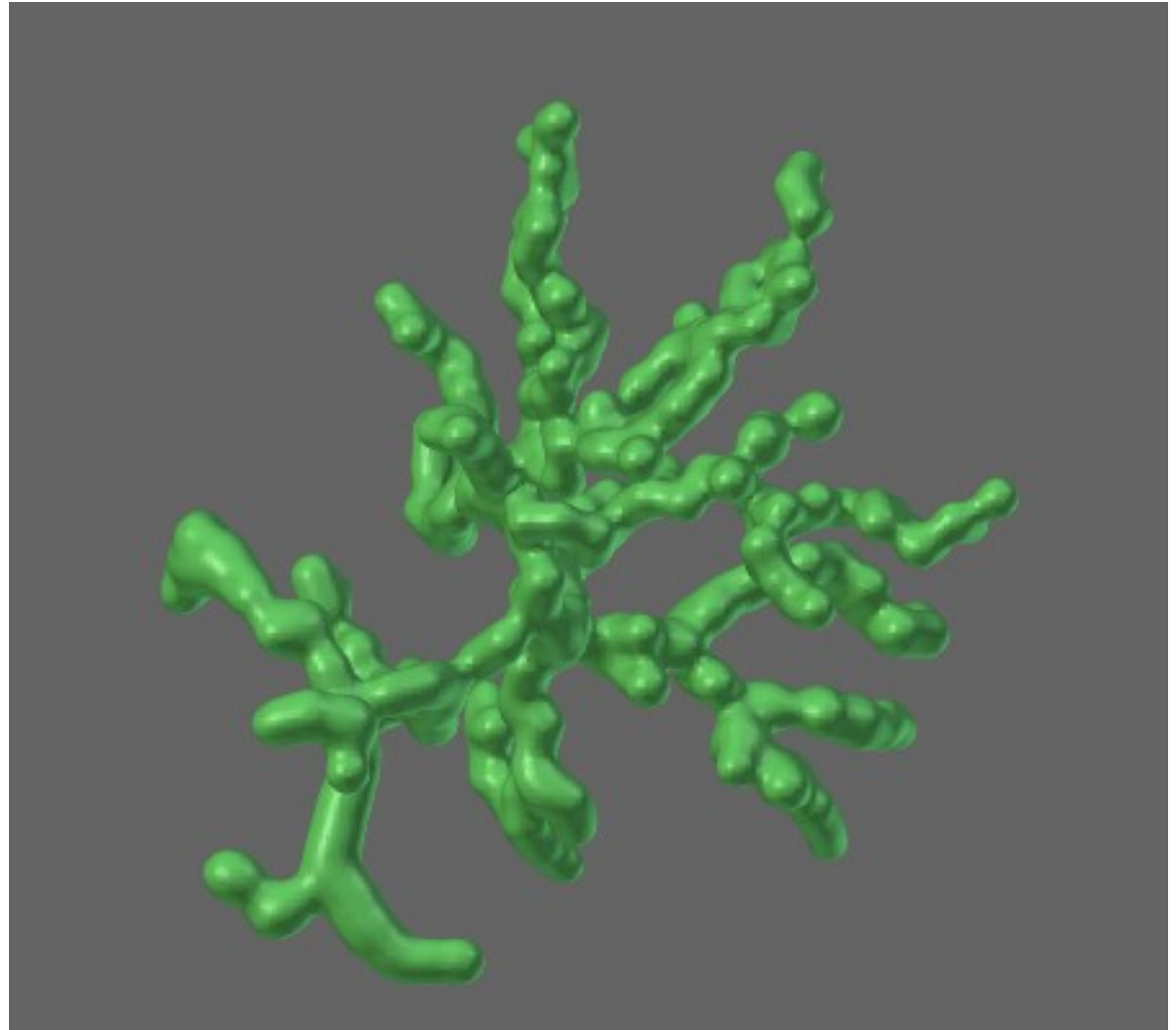
Mesh Modeling

3D reconstruction

Lumen

3) “Shrink-wrap”
the spheres
together.

Constructive Solid
Geometry.
Octree remesh.
STL files.

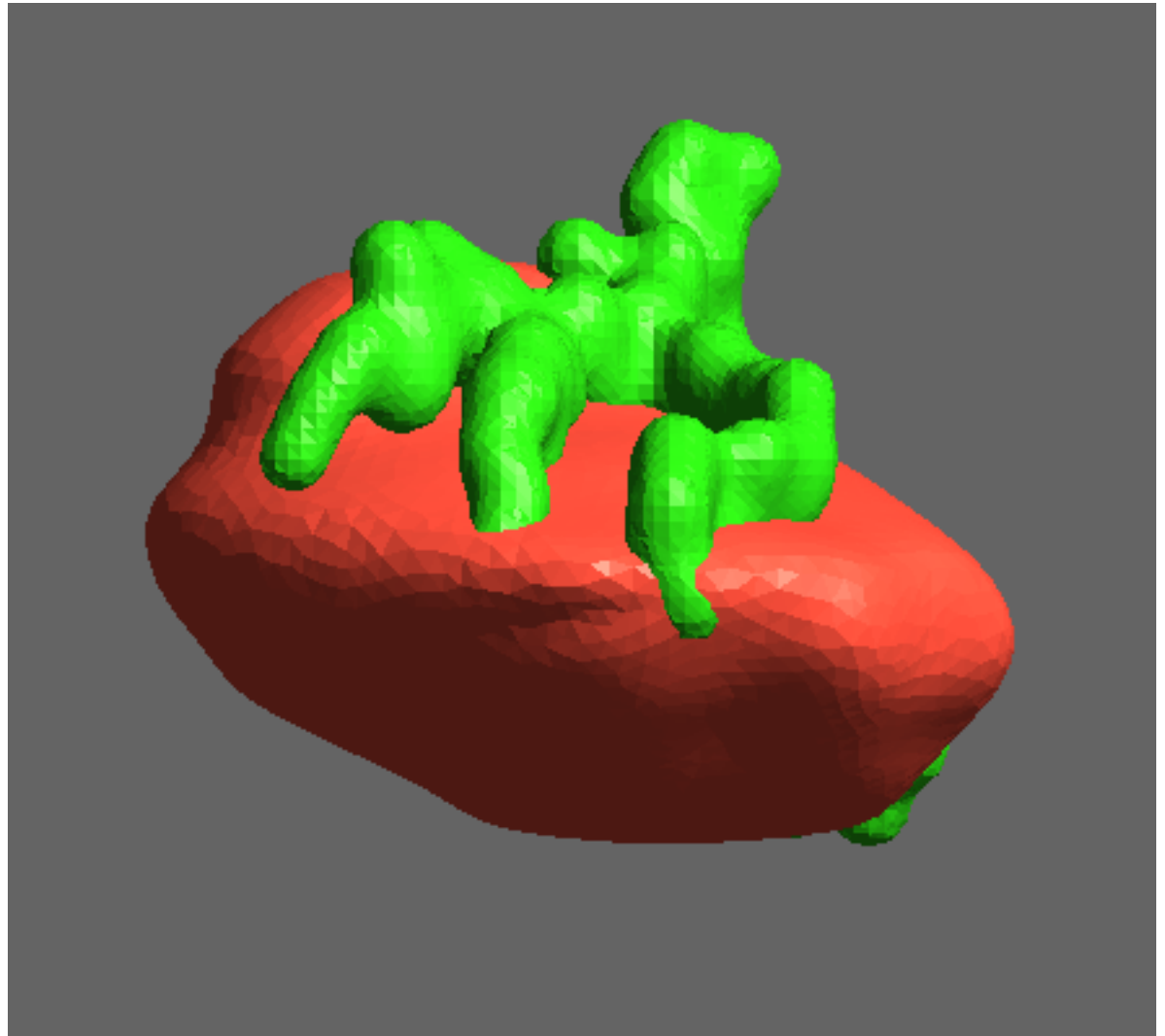


Reconstructed 3D Model

Cells & Lumen

Each cell is held
by a lumen “claw”.

The lumen has a
tree-like branching
structure.

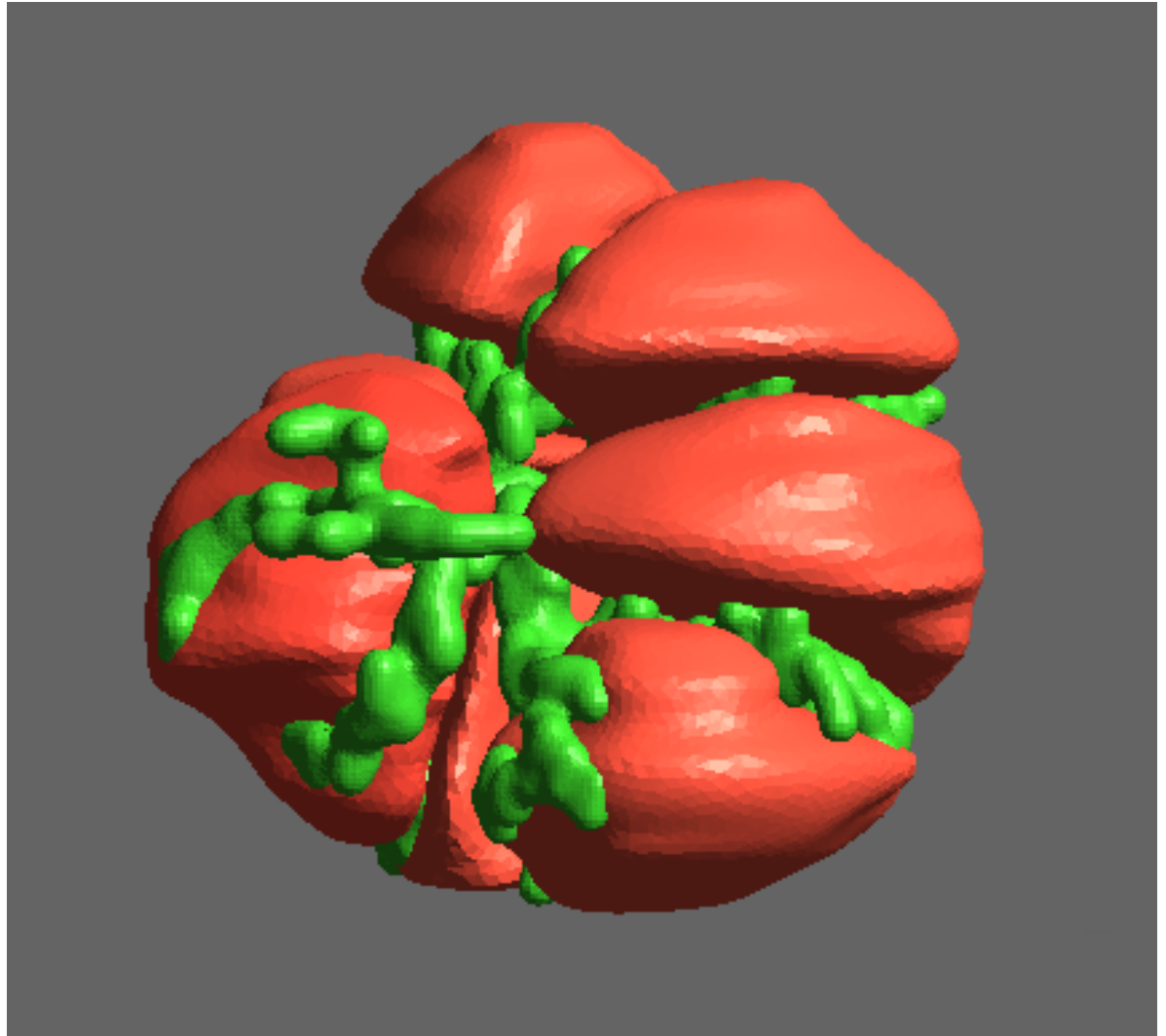


Reconstructed 3D Model

Cells & Lumen

The cells are grouped in tight clusters.

Each lumen has a central trunk.

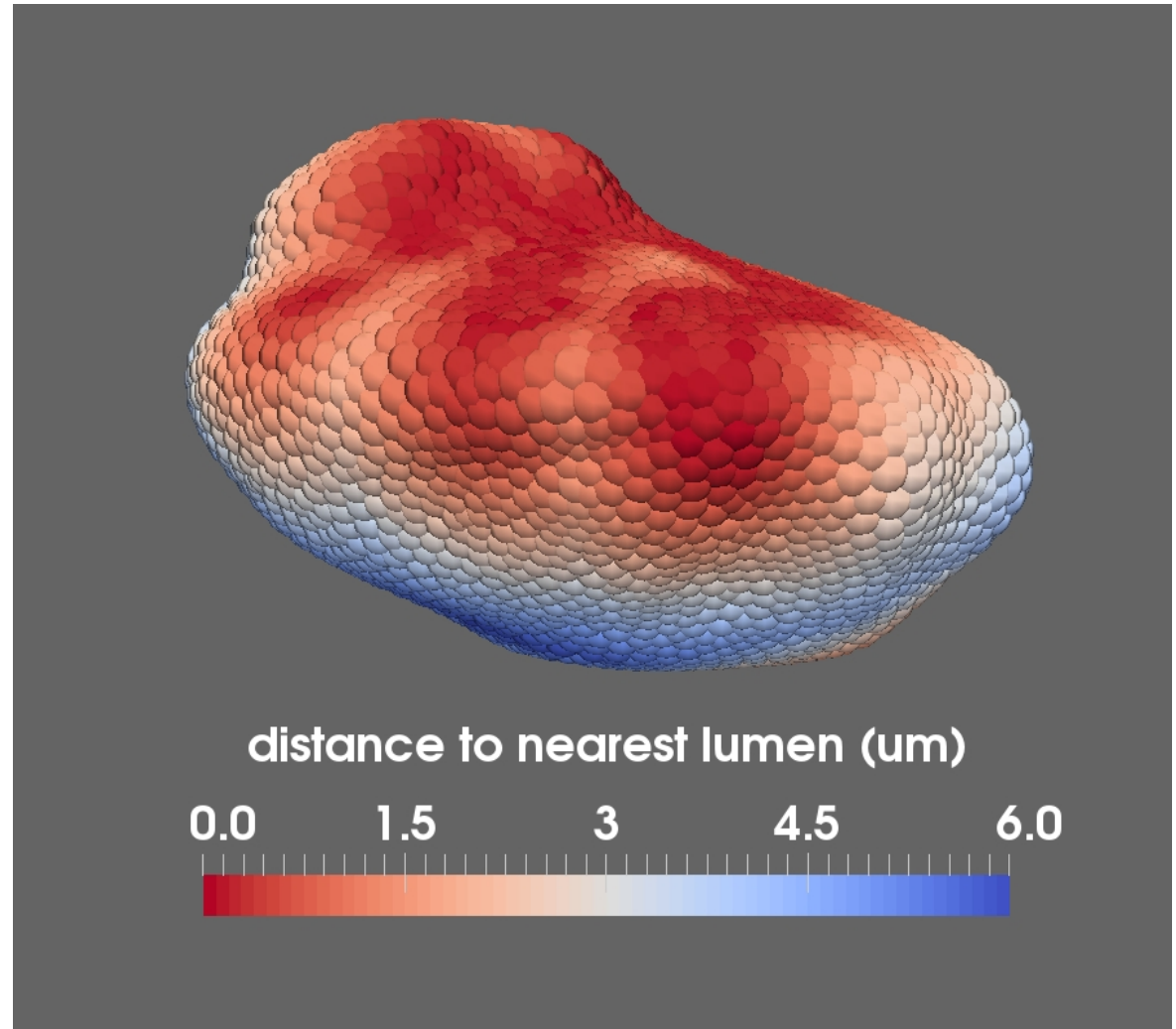


Solid Volumetric Mesh

Tetrahedral mesh for finite element simulation

Nodal view

Imprint of the luminal
“claw”.

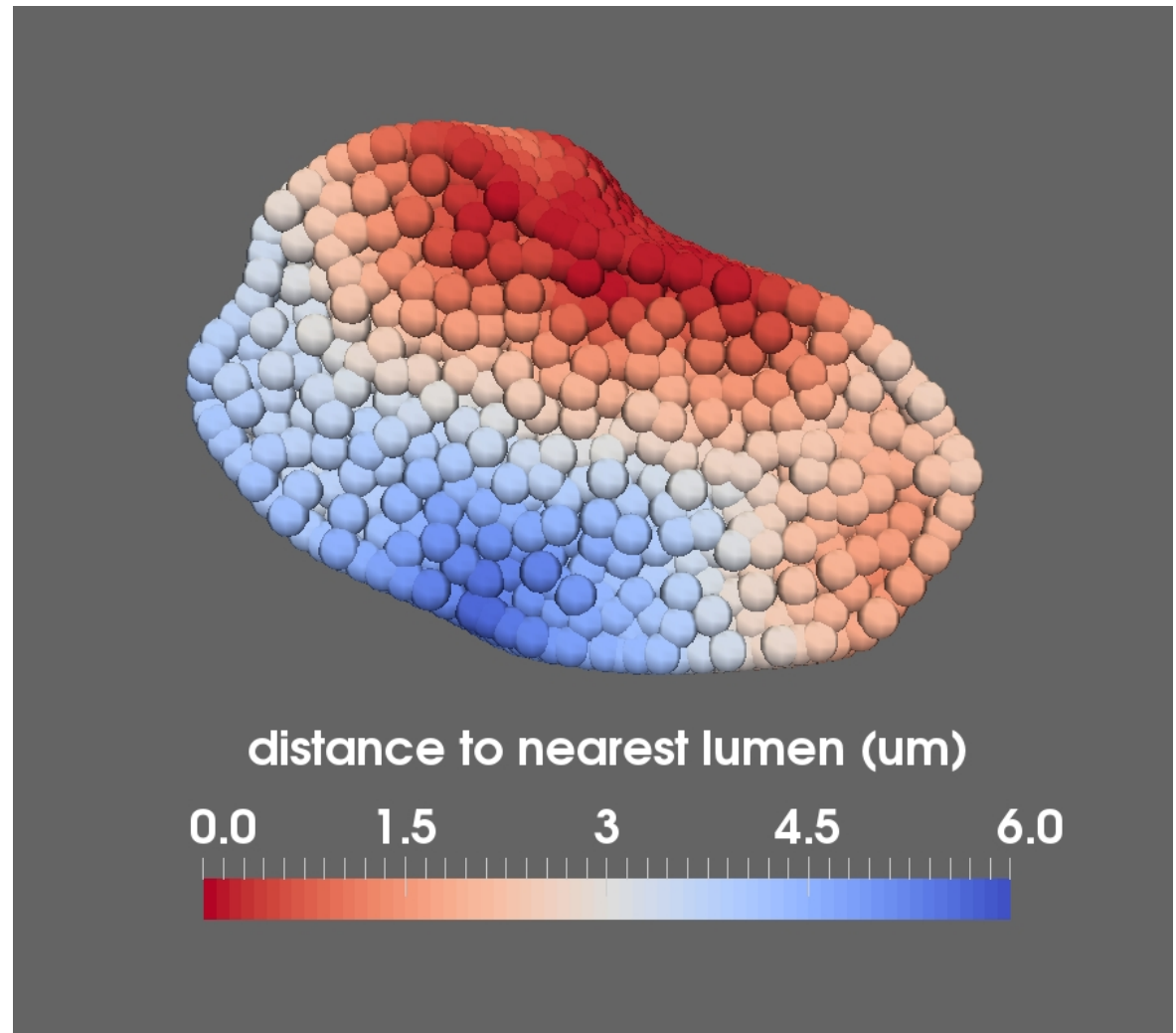


Solid Volumetric Mesh

Tetrahedral mesh for finite element simulation

Nodal view
(cut-away)

Imprint of the luminal
“claw”.



FEM Simulation

Calcium waves

Partial differential equations model the cell calcium dynamics.

Reaction-Diffusion

$$\frac{\partial c}{\partial t} = D_c \nabla^2 c + (J_{\text{IPR}} + J_{\text{leak}})(c_e - c) - J_{\text{serca}}$$

$$\frac{\partial p}{\partial t} = D_p \nabla^2 p + V_{\text{PLC}}(\vec{x}) - V_{\text{deg}} \left(\frac{c^2}{K_{3K}^2 + c^2} \right) p$$

$$\frac{\partial h}{\partial t} = \frac{h_\infty - h}{\tau}$$

$$J_{\text{serca}} = V_s \frac{c^2}{K_s^2 + c^2}$$

$$J_{\text{IPR}} = k_{\text{IPR}}(\vec{x}) P_O$$

$$P_O = \phi_c \phi_p h$$

$$\phi_c = \frac{c^3}{K_a^3 + c^3}$$

$$\phi_p = \frac{p^4}{K_p^4 + p^4}$$

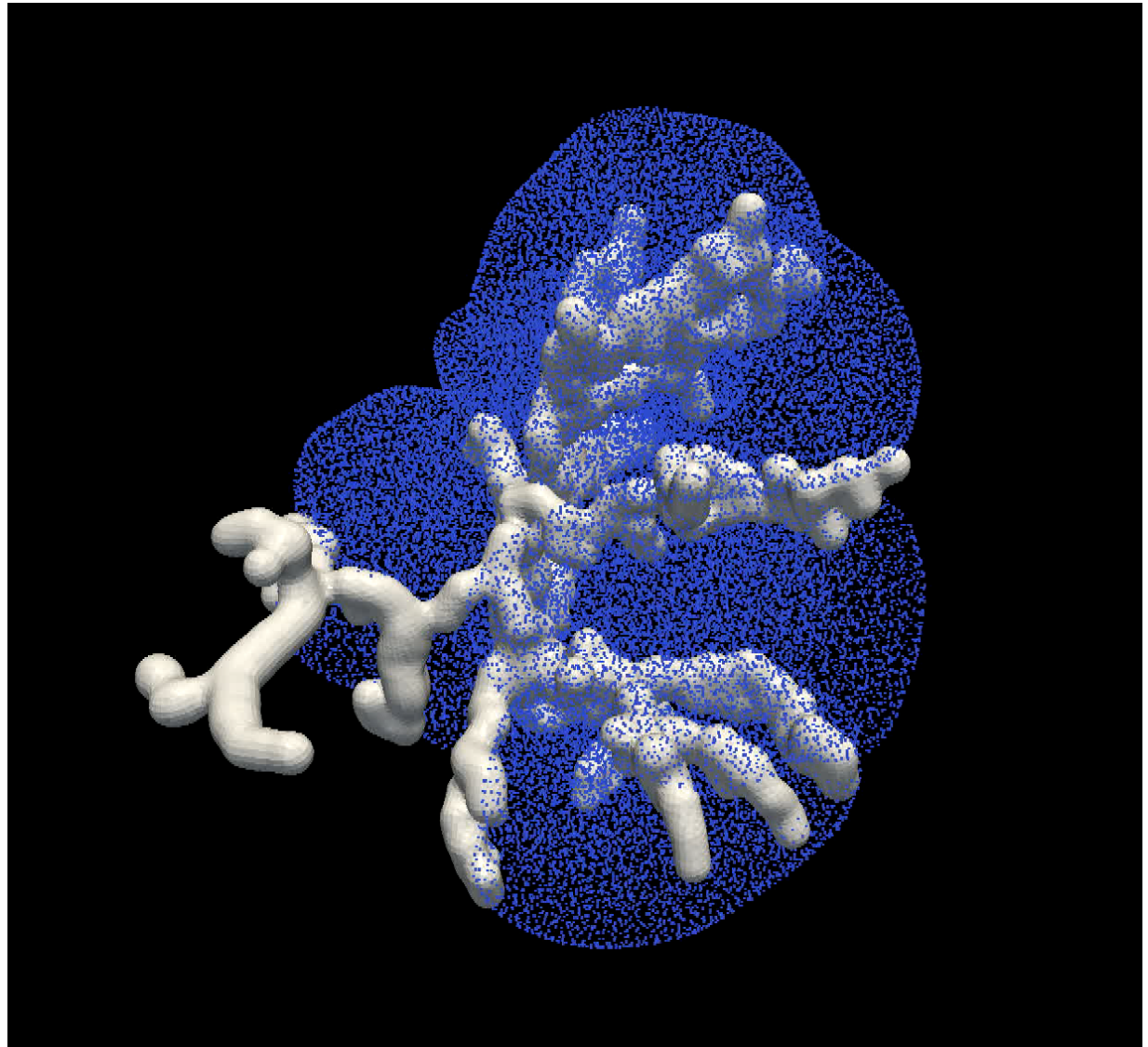
$$h_\infty = \frac{K_i^2}{K_i^2 + c^2}$$

$$c_e = (c_t - c)/\gamma$$

FEM Simulation Results

Calcium waves

Wave-fronts propagate between the apical and basal ends of each cell.

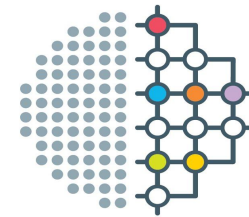
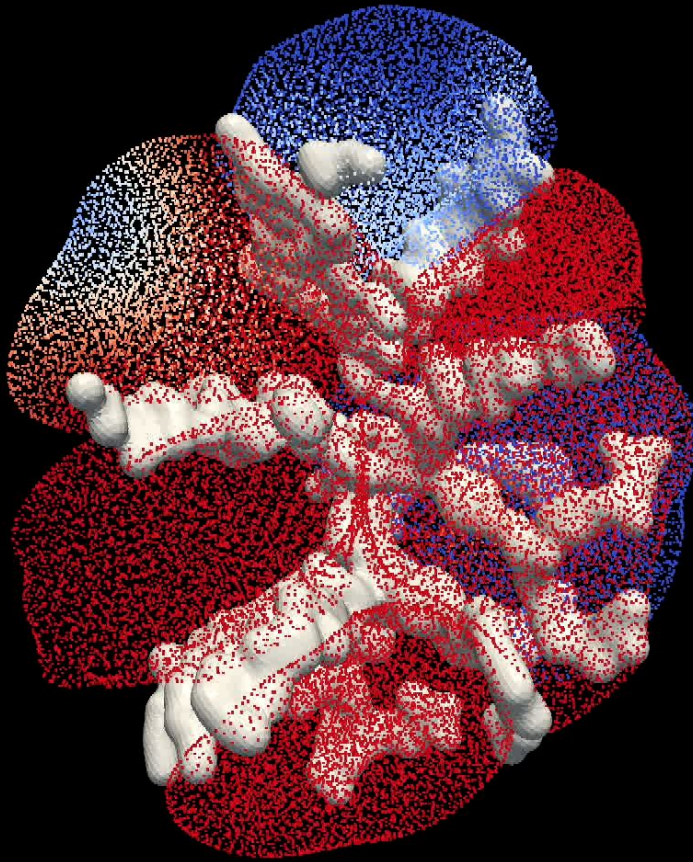


What's next?

- Model and simulate interaction between cells (i.e. gap junctions)
- Include fluid flow in lumen (computational fluid dynamics)
- Scale up to a larger number of cells

More to come...

Questions & Answers?



NeSI
New Zealand eScience
Infrastructure

www.nesi.org.nz